5'-**GGTGGTTTGTTTG**-3' (SEQ ID NO:5), LNA nucleosides in bold) was mixed with T4 polynucleotide Kinase (5 Units; New England Biolabs) and 6 μl γ-^{32P}ATP (3000 Ci/mmol, Amersham) in a buffer containing 70 mM Tris-HCl (pH 7.6), 10 mM $MgCl_2$, 5 mM dithiotretiol (final volume 20 μl).

At page 87, please replace the sentence starting at line 12 and ends at line 17, with:

The sequence and extent of LNA modification were as follows (where LNA monomers are in bold):

Control 5' GGT GGT TTG TTT G 3' (SEQ ID NO:6)

(1) 5' GGT GGT **T**TG TTT G 3' (SEQ ID NO:7)

(2) 5' GGT GG**T TTG** TTT G 3' (SEQ ID NO:8)

(3) 5' **GGT GGT TTG TTT** G 3' (SEQ ID NO:9)

At page 87, please replace the sentence that starts at line 32, and ends at page 88, line 4 with:

The following 15mer primers and a mixture of 8 to 32 base oligonucleotide markers were 5' end labelled with [γ ^{33}P] ATP and T4 polynucleotide kinase (where LNA monomers are in bold):

P1 5'-TGC ATG TGC TGG AGA-3' (SEQ ID NO:10)

P2 5'- GC ATG TGC TGG AGA T-3' (SEQ ID NO:11)

PZ1 5'-**T**GC ATG **T**GC **T**GG AGA-3' (SEQ ID NO:12)

PZ2 5'- GC ATG TGC TGG AGA **T**-3' (SEQ ID NO:13)

At page 88, please replace the sentence starting at line 21, at ends at line 23, with:

A 15mer primer (sequence: 5'-TGC ATG TGC TGG AGA-3' (SEQ ID NO:14)) and a mixture of 8 to 32 base oligonucleotide markers were 5' end labelled with [γ ^{33}P) ATP and T4 polynucleotide kinase.

At page 89, please replace the sentence starting at line 14, and ends at line 20, with:

The sequences of the primer and templates are (LNA monomer in bold):

Primer 5′ TGCATGTGCTGGAGA 3′ (SEQ ID NO: 15)

Template 1 3′ **ACGTACACGACCTCT**ACCTTGCTA 5′ (SEQ ID NO:16)

Template 2 3′ **ACGTACACGACCTCT**CTTGATCAG 5′ (SEQ ID NO:17)

Template 3 3′ **ACGTACACGACCTCT**TGGCTAGTC 5′ (SEQ ID NO:18)

Template 4 3′ **ACGTACACGACCTCT**GAACTAGTC 5′ (SEQ ID NO:19)

At page 91, please replace the sentence starting at line 11, and ends at line18, with:

The following 15mer primers and 8 to 32 base oligonucleotide markers were 5′ end labelled with [γ ^{33}P] ATP and T4 polynucleotide kinase (LNA monomer in bold):

P2 5′- GC ATG TGC TGG AGA T-3′ (SEQ ID NO:20)

PZ2 5′- GC ATG TGC TGG AGA T-3′ (SEQ ID NO:21)

Reactions were boiled for 5 min after labelling to remove any PNK activity. 8 picomoles of each primer was hybridised to 25 pmoles Template (sequence: 3′- ACG TAC ACG ACC TCT ACC TTG CTA-5′ (SEQ ID NO:22)) in x2 Klenow buffer.

At page 92, please replace the paragraph that starts at line 8 and ends at line 16, with:

PCR reaction mixture for Amplicon 1

1 μl pUC19 (1 ng/μl),

1 μl reverse primer (5′-AACAGCTATGACCATG-3′) (SEQ ID NO:23) (20 μM),

1 μl forward primer (5′- GTAAAACGACGGCCAGT-3′) (SEQ ID NO:24) (20 μM),

10 μl dUTP-mix (2 mM dATP, 2 mM dCTP, 2 mM dGTP and 6mM dUTP), 1.5 μl DIG-11-dUTP (1 mM)

10 μl 10x Taq buffer (Boehringer Mannheim incl $MgCl_2$)

G8 Cont

1 μl Taq polymerase (Boehringer Mannheim) 5 U/μl
H_2O ad 100 μl--

At page 92, please replace the paragraph at page 92, lines 18-26, with:

PCR reaction mixture for Amplicon 2
1 μl pUC19 (1 ng/μl),
0.4 μl primer 3 (5'-GATAGGTGCCTCACTGAT-3') (SEQ ID NO:25) (50 μM),
0.4 μl primer 4 (5'-GTCGTTCGCTCCAAGCTG-3') (SEQ ID NO:26) (50 μM),
G9
10 μl dUTP-mix (2 mM dATP, 2 mM dCTP, 2 mM dGTP and 6mM dUTP),
1.5 μl DIG-11-dUTP (1 mM)
10 μl 10x Taq buffer (Boehringer Mannheim incl $MgCl_2$)
1 μl Taq polymerase (Boehringer Mannheim) 5 U/μl
H_2O ad 100 μl

At page 93, please replace the sentence starting at line 3, and ends at line 9, with:

The following capture probes were used: B-DNA1 (biotin-ATGCCTGCAGGTCGAC-3' SEQ ID NO:27); DNA probe specific for amplicon 1), B-DNA2 (biotin-GGTGGTTTGTTTG-3' SEQ ID NO:28); DNA probe specific for amplicon 2) and B-LNA2 (biotin--
G10
GGTGGTTTGTTTG-3' (SEQ ID NO:29), LNA nucleosides in bold; LNA probe specific for amplicon 2). Reactions were heated to 95°C for 5 min in order to denature amplicons and allowed to cool at 25°C for 15 min to facilitate hybridisation between the probe and the target amplicon strand.

At page 94, please replace the sentence starting at line 3 and ends at line 6, with:

Two identical sets of 10 µl reactions of amplicon 1 or 2 (prepared as in Example 143) were mixed with either 1, 5 or 25 pmol of the B-LNA2 capture probe (biotin-
G11
GGTGGTTTGTTTG-3' (SEQ ID NO:30), LNA nucleosides in bold; probe specific for

G11 Cont amplicon 2) in 1 x SSC (0.15 M NaCl, 15mM citrate, pH 7.0) in a total volume of 450 μl.--

At page 95, please replace the sentence that starts at line 6, and ends at line 11, with:

Wells of a streptavidin coated microtiter plate (Boehringer Mannheim) were incubated for 1 hour with either 5 pmol of the B-DNA2 probe (biotin-GGTGGTTTGTTTG-3′ SEQ ID NO:31); DNA probe specific for amplicon 2) or the B-LNA2 probe (biotin-**GGTGGTTTGTTT**G-3′ (SEQ ID NO:32), LNA nucleosides in bold; LNA probe specific for amplicon 2) in a total volume of 100 μl 1xSSC (0.15 M NaCl, 15mM citrate, pH 7.0).

G12

At page 96, please replace the sentence that starts at line 9, and ends at line 20, with:

--Three DIG labelled amplicons from Nras sequence (ref.: Nucleic Acid Research, **1985,** Vol. 13, No. 14, p 52-55) were generated by PCR amplification as follows:
PCR primers:
Forward primer: 5′-CCAGCTCTCAGTAGTTTAGTACA-3′ SEQ ID NO:33) bases 701-723 according to the NAR reference.
910 by reverse primer: 5′-GTAGAGCTTTCTGGTATGACACA-3′ (SEQ ID NO:34) bases 1612-1590 (reverse sequence according to NAR ref.).
600 by reverse primer: 5′-TAAGTCACAGACGTATCTCAGAC-3′ (SEQ ID NO:35) bases 1331-1308 (reverse sequence according to NAR ref.).
200 by reverse primer: 5′-CTCTGTTTCAGACATGAACTGCT-3′ (SEQ ID NO:36) bases 909-886 (reverse sequence according to NAR ref.).

G13

At page 96, please replace the sentence that starts at line 32, and ends at page 97, line 5, with:

Assay conditions: Wells of a streptavidin coated micro-titer plate (Boehringer Mannheim; binding capacity of 20 pmol biotin per well) were incubated for 1 hour in 5 x SSCT (0.75 M NaCl, 75 mM citrate, pH 7.0, 0.1 % Tween 20) at 37°C with either 1 pmol of DNA Nras

G14

Cap A (biotin-5'-TTCCACAGCACAA-3') (SEQ ID NO:37), LNA/DNA Nras Cap A (biotin-5'-TTCCA**CAG**CACAA-3') (SEQ ID NO:38, LNA Nras Cap A (biotin-5'-**TTCCACAGCACA**A-3') (SEQ ID NO:39), DNA Nras Cap B (biotin-5'-AGAGCCGATAACA-3') (SEQ ID NO:40), LNA/DNA Nras Cap B (biotin-5'-AGAGC**CGAT**AACA-3') (SEQ ID NO:41) or LNA Nras Cap B (biotin-5'-**AGAGCCGATAAC**A-3') (SEQ ID NO:42); LNA nucleosides in bold.

G14 Cont

At page 98, please replace the sentence starting at line 3, and ends at line 6 with:

The ability of an LNA modified oligo (5'-GGTGG**TTTG**TTTG-3') (SEQ ID NO:43), LNA nucleosides in bold) to serve as primer in template dependent, enzymatic elongation were investigated with 3 different classes of polymerases.

G15

At page 98, please replace the sentences that start at line 9, and end at line 13 with the following:

As control the extension reactions were conducted using the identical unmodified DNA primer (5'-GGTGGTTTGTTTG-3') (SEQ ID NO:44). The LNA and DNA primers were labelled with ^{32}P-γ-ATP as previously described in Example 137. A 50mer DNA oligo (5'-AAAAATCGACGCTCAAGTCAGAAAAGCATCTCACAAACAAACAAACCACC-3') (SEQ ID NO:45) was used as template.

G16

At page 99, please replace the sentences that starts at line 10, and end at line 19, with the following:

The ability of LNA modified oligos to act as primers in PCR amplification was analysed with three oligos differing only in the number of LNA nucleosides they contained: 4 LNA nucleosides (AL2 primer: 5'-GGTGG**TTTG**TTTG-3' (SEQ ID NO:46), LNA nucleosides in bold), 1 LNA nucleoside (AL10 primer: 5'-GGTGGTTTGTTTG-3' (SEQ ID NO:47), LNA nucleoside in bold) and no LNA nucleoside (FP2 primer: 5'-GGTGGTTTGTTTG-3') (SEQ ID NO:48. The PCR reactions (100 µl) contained either no template (control), 0.01 ng, 0.1 ng or 1

G17

G17 Cont

ng of template (pUC19 plasmid), 0.2 μM reverse primer (5'-GTGGTTCGCTCCAAGCTG-3') (SEQ ID NO:49), 0.2 μM of either the AL2, AL10 or FP2 forward primer, 200 μM of dATP, dGTP, dCTP and dTTP, 10 mM Tris-HCl pH 8.3, 1.5 mM $MgCl_2$, 50 mM KCl and 2.5U of the BM-Taq polymerase.

At page 100, please replace the sentence that starts at line 26, and ends at line 29, with the following:

G18

Either 25 pmol/μl or 12.5 pmol/μl of an anthraquinone DNA oligo (5'-AQ-CAG CAG TCG ACA GAG-3') (SEQ ID NO:50) or an anthraquinone LNA modified DNA oligo (5'-AQ-CAG CAG TCG ACA GAG-3' (SEQ ID NO:51); LNA monomer is underlined) was spotted (1 μl/spot) in 0.2 M LiCl on a polycarbonate slide (Nunc). The oligos were irradiated for 15 min with soft UV light.

At page 100, please replace the sentence that starts at line 30, and ends at line 34, with the following:

G19

After irradiation the slide was washed three times in Milli-Q water and air-dried. 25 ml of 0.5 pmol/μl of complimentary biotinylated oligomer (5'-biotin- CTC TGT CGA CTG CTG-3') (SEQ ID NO:52) was hybridised to the immobilised oligomers in 5 x SSCT (75 mM Citrate, 0.75 M NaCl, pH 7.0, 0.1 % Tween 20) at 50°C for 2 hours.

At page 107, please replace the sentence that starts at line 23, and ends at line 27 with the following:

G20

The 15mer primer (sequence: 5'-TGC ATG TGC TGG AGA-3') (SEQ ID NO:53) was used to prime the following short oligonucleotide sequences (LNA monomer in bold):

Template 1 3'-ACG TAC ACG ACC TCT ACC TTG CTA-5' (SEQ ID NO:54)

Template TZ1 3'-ACG TAC ACG ACC TCT ACC **T**TG CTA-5' (SEQ ID NO:55)

At page 109, please replace the sentence that starts at line 9, and ends at line 15 with the following:

10 pmol of a oligodeoxynucleotide (ODN) (ODN#10: 5'-TTA ACG TAG GTG CTG GAC TTG TCG CTG TTG TAC TT-3' (SEQ ID NO:56), a 35-mer complementary to human Cathepsin D) and 10 pmoles of two LNA oligos: AL16 (5'-d(**TGT GTG AAA TTG TTA** T)-3' (SEQ ID NO:57), LNA nucleosides in bold) and AL17 (5'-d(**ATA AAG TGT AAA** G)-3' (SEQ ID NO:58), LNA nucleosides in bold) were mixed with T4 polynucleotide Kinase (10 units, BRL cat. no. 510-8004SA), 5 µl gamma $^{-32}$P-ATP 5000 Ci/mmol, 10 uCi/µl (Amersham) in kinase buffer (50 mM Tris/HCl pH 7,6, 10 mM $MgCl_2$, 5 mM DTT, 0.1 mM EDTA).

At page 111, please replace the Table entitled "Oligonucleotides tested" located at the top of page 111, with:

Table: Oligonucleotides tested

Name	Sequence (LNA monomers in bold)	Characteristics
AL16	5'-**TGT GTG AAA TTG TTA** T-3' (SEQ ID NO: 59)	LNA, enzym. FITC labeled
AL17	5'-**ATA AAG TGT A AA** G-3' (SEQ ID NO:60)	LNA, enzym. FITC labeled
EQ3009-01	5'-**TGC CTG CAG GTC GAC** T-3' (SEQ ID NO:61)	LNA-FITC-labeled
EQ3008-01	5'-TGC CTG CAG GTC GAC T-3' (SEQ ID NO:62)	DNA-FITC-labeled--

At page 112, please replace the sentence starting at line 34, and ends at page 113, line 3, with the following:

G23 Two LNA oligos: AL16 (5'-**TGT GTG AAA TTG TTA** T-3' SEQ ID NO: 63), LNA nucleosides in bold) and AL17 (5'-**ATA AAG TGT AAA** G-3' SEQ ID NO:64), LNA nucleosides in bold) were labeled with fluorescein as described in Example 128. MCF-7 human breast cancer cells were cultured as described in Example 154.

At page 115, please replace the sentence that starts at line 18, and ends at line 24, with the following:

The following poly dT primers were tested (LNA monomers are in bold):

G24
RTZ1 5'-TTT TTT **T**TT TTT TT-3' (SEQ ID NO:65)
RTZ2 5'-TTT TTT **TT**T TTT TT-3' (SEQ ID NO:66)
RTZ3 5'-TTT TT**T TTT** TTT TT-3' (SEQ ID NO:67)
RTZ4 5'-TTT T**TT TTT** T**T**T TT-3' (SEQ ID NO:68)
RTZ5 5'-**TTT TTT TTT** T-3' (SEQ ID NO:69)

At page 116, please replace the sentence that starts at line 24, and ends at line 28, with the following:

Three oligos were synthesised by chemistry (Amy Mueller) for evaluation in poly (rA) binding.

G25
- NH_2(T8)-T Control
- NH_2(T15)-T Control SEQ ID NO:70)
- NH_2(LNA-**T**8)-T Test-

Please replace Table 1 (cont.) (3), with the following:

--Table 1 (cont.)

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
5'-d(GGTGGTTTGTTTG)-3' (SEQ ID NO:71)					
	5'-d(CAAACAAACCACA)-3' (SEQ ID NO:72)	39	31	47	55
	5'-(CAAACAAACCACA)-3' (SEQ ID NO:73)	39A	32	52	
5'-d(GGTGG**TTTG**TTTG)-3' (SEQ ID NO:74)					
	5'-d(CAAACAAACCACA)-3' (SEQ ID NO:75)	40	40	57	67
	5'-(CAAACAAACCACA)-3' (SEQ ID NO:76)	40A	50	70	
d(**GGTGGTTTGTTT**G)-3' (SEQ ID NO:77)					
	5'-d(CAAACAAACCACA)-3' (SEQ ID NO:78)	41	67	83	>90
	5'-(CAAACAAACCACA)-3' (SEQ ID NO:79)	41A	85	>93	
5'-d(TTTTTTTTTTTTTT)-3' (SEQ ID NO:80)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:81)	42		36	
	5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:82)	43		32	
5'-d(TTTTTT**T**TTTTTTT)-3' (SEQ ID NO:83)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:84)	44		36	
	5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:85)	45		32	
5'-d(TTTTTT**TT**TTTTTT)-3' (SEQ ID NO: 86)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID	46		34	

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
NO:87)					
5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:88)		47		40	
5'-d(TTTTT**TTTT**TTTTT)-3' (SEQ ID NO:89)					
5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:90)		48		42	
5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:91)		49		52	
5'-d(TTTTTTTTTTTTTT)-3' (SEQ ID NO:92)					
5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:93)		50		47	
5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:94)		51		53	
5'-d(**TTTTTTTTT**T)-3' (SEQ ID NO:95)					
5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:96)		52		80	
5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:97)		53		70	
5'-d(AAAACAAAA)-3'		54		63	
5'-d(AAAAGAAAA)-3'		55		55	
5'-d(AAAATAAAA)-3'		56		65	
5'-d(G**T**GA**A**A**T**GC)-3'					
5'-d(GCATATCAC)-3'		57		26	
5'-d(GCATTTCAC)-3'		58		45	
5'-d(GCATGTCAC)-3'		59		23	
5'-d(GCATCTCAC)-3'		60		25	
5'-d(GTGA^{Me}CATGC)-3'					
5'-d(GCATATCAC)-3'		61		<15	
5'-d(G**T**GA^{Me}C**A**T**G**C)-3'					

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
5'-d(GCATATCAC)-3'		63		32	
5'-d(GCATTTCAC)-3'		64		27	
5'-d(GCATGTCAC)-3'		65		53	
5'-d(GCATCTCAC)-3'		66		32	

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Please replace Table 2 (Monomer V) (6), with the following:

--Table 2 Monomer V

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
5'-d(TTTTTTTTTTTTTTT)-3' (SEQ ID NO:98)					
	5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:99)			32	
	5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:100			27	
5'-d(TTTTTTTTTTTTTTT)-3' (SEQ ID NO:101)					
	5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:102)			31	
	5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:103)			28	
5'-d(TTTTTTTTTTTTTTT)-3' (SEQ ID NO:104)					
	5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:105)			30	
	5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:106)			23	
5'-d(TTTTTTTTTTTTTTT)-3' (SEQ ID NO:107)					
	5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:108)			23	
	5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:109)			31	
5'-d(TTTTTTTTTTTTTTT)-3' (SEQ ID NO:110)					
	5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:111)			23	
	5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:112)			16	
5'-d(TTTTTTTTTTTTTT)-3' (SEQ ID NO:113)					
	5'-d(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:114)			<10	
	5'-(AAAAAAAAAAAAAAA)-3' (SEQ ID NO:115)			42	
	5'-(AAAAAAGAAAAAAA)-3' (SEQ ID NO:116)			37	
5'-d(GTGATATGC)-3'					
	5'-d(GCATATCAC)-3'			26	

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
5'-(GCAUAUCAC)-3'				27	

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Please replace Table 3 (Monomer X) (7), with the following:

--Table 3 Monomer X

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
5'-d(TTTTTTT**T**TTTTTT)-3' (SEQ ID NO:117)					
5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:118)				23	
5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:119)				23	
5'-d(TTTTTT**TT**TTTTTT)-3' (SEQ ID NO:120)					
5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:121)				19	
5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:122)				23	
5'-d(TTTTTTT**TT**TTTTT)-3' (SEQ ID NO:123)					
5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:124)				9	
5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:125)				15	
5'-d(TTTTT**TTT**TTTTT)-3' (SEQ ID NO:126)					
5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:127)				5	
5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:128)				14	

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Please replace Table 4 (Monomer Y) (8), with the following:

--Table 4 Monomer Y

Oligo	Target	T_m No.	T_m (°C) Na_2HPO_4/EDTA	T_m (°C) Na_2HPO_4/NaCl/EDTA	T_m (°C) Na_2HPO_4/TMAC
5'-d(TTTTTTT**T**TTTTTT)-3' SEQ ID NO:129)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:130)			36	
	5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:131)			37	
5'-d(TTTTTTT**TT**TTTTTT)-3' (SEQ ID NO:132)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:133)			35	
	5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:134)			37	
5'-d(TTTTTTT**TT**TTTTTT)-3' (SEQ ID NO:135)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:136)			35	
	5'-(AAAA.AAAAAAAAAA)-3' (SEQ ID NO:137)			36	
5'-d(TTTTT**TTTT**TTTTT)-3' (SEQ ID NO:138)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:139)			32	
	5'-(AAAA.AAAAAAAAAA)-3' (SEQ ID NO:140)			33	
5'-d(TTTT**T**TTT**T**TTT**T**TT)-3' (SEQ ID NO:141)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:142)			36	
	5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:143)			36	
5'-d(**TTTTTTTTTTTTT**T)-3' (SEQ ID NO:144)					
	5'-d(AAAAAAAAAAAAAA)-3' (SEQ ID NO:145)			58	
	5'-(AAAAAAAAAAAAAA)-3' (SEQ ID NO:146)			58	

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